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STUDIES ON CEPHALOSPORIN ANTIBIOTICS

II. SYNTHESIS, ANTIBACTERIAL ACTIVITY AND ORAL ABSORPTION OF 3-ALKOXYCARBONYLMETHOXY-7β-[(Z)-2-(2-AMINOTHIAZOL-4-YL)-2-(0-SUBSTITUTED OXYIMINO)-ACETAMIDO]CEPHALOSPORINS

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The synthesis, antibacterial activity and oral absorption in rats of 3-alkoxycarbonylmethoxy-7 β -[(Z)-2-(2-aminothiazol-4-yl)-2-(O-substituted oxyimino)acetamido]cephalosporins (1) are described. In this cephalosporin series, 7β -[(Z)-2-(2-aminothiazol-4-yl)-2-(carboxymethoxyimino)acetamido]cephalosporins (1b, 1i and 1j) with a lower alkoxycarbonylmethoxy group at the C-3 position of a cephem nucleus exhibited not only potent activity against Gram-negative bacteria but also good oral absorption in rats. Structure-activity relationships of 1 are also presented.

During the past decade, cephalosporins^{1~5)} bearing an 2-aminothiazole-oxime moiety at the C-7 position of a cephem nucleus have been developed successively. They have excellent antibacterial activities against Gram-positive and Gram-negative bacteria including β -lactamase producing strains. However, all of the new cephalosporins clinically used are not suitable for oral administration because of their low absorption from the gastro-intestinal tract. As orally active cephalosporins, only cephalexin⁶⁾ and its analogues with D-phenylglycine or closely related moieties at the C-7 position are now available in clinical use. But their activities against Gram-negative bacteria are much lower than those of the 2-aminothiazole-oxime type cephalosporins.

Therefore, much research has been undertaken aiming at obtaining a new orally active cephalosporin possessing the same antibacterial properties as those of the new parenteral cephalosporins. Recently, some orally active 2-aminothiazole-oxime type cephalosporins such as cefixime (FK027)⁷) and T-2588⁸ have been reported.

In the course of our research on new cephalexin analogues, we have found an interesting fact that the analogues bearing an alkoxycarbonylmethoxy group at the C-3 position exhibit much higher peak serum levels than that of cephalexin after oral administration to rats⁹. Then, we became interested

in the application of the novel C-3 alkoxycarbonylmethoxy substituents into the 2-aminothiazole-oxime type cephalosporins as an attempt to improve their oral absorption. This paper describes the synthesis, antibacterial activity and oral absorption in rats of new cephalosporins (1) as shown in Fig. 1.





Chemistry

The new cephalosporin derivatives $(1a \sim 11)$ listed in Tables 1 and 2 were prepared by the methods shown in Scheme 1. Diphenylmethyl (Bh) 7 β -phenoxyacetamido-3-hydroxycephalosporinate 1-oxide $(2)^{10}$ was reacted with 2-bromoacetic acid ester derivatives (3) in the presence of *N*,*N*-diisopropylethylamine to afford C-3 *O*-substituted cephem compounds (4). Reduction of the sulfoxide of 4 using phosphorus tribromide (PBr₃) in *N*,*N*-dimethylformamide gave the corresponding sulfides (5). Then, the C-7 phenoxyacetyl side chain was cleaved by the known imino-chloride method¹¹⁾ to yield Bh 7 β -aminocephalosporinates (6). The coupling reaction of (*Z*)-2-(2-tritylaminothiazol-4-yl)-2-(*O*-substituted oxyimino)acetic acid derivatives (7) with 6 was carried out *via* their acid chlorides (formed with phosphorus pentachloride) in dichloromethane at low temperature to give the protected cephalosporins (8). The protecting groups in 8 were removed with trifluoroacetic acid and anisole to afford the desired compounds (1).

Antibacterial Activity and Oral Absorption

The *in vitro* antibacterial activities of the new cephalosporins (1) against selected Gram-positive and Gram-negative bacteria and their peak serum levels as a measure of gastro-intestinal absorption after oral administration (50 mg/kg) to rats are summarized in Tables 1 and 2. For comparison, the MIC values and the peak serum levels of cephalexin and cefixime are listed at the bottom of Tables 1 and 2.

Table 1 shows the effect of the oxime O-substituent (R_1) on antibacterial activity and oral absorption. Against the Gram-negative bacteria, all of the derivatives $(1a \sim 1g)$ exhibited high activity compared with cephalexin. These compounds except 1f showed the activity comparable to that of cefixime. However, their activity against *Staphylococcus aureus* was lower than that of cephalexin





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	Compound		MIC (µ	ug/ml, 10 ⁶ cl	fu/ml)ª		Peak serum level - (ug/ml) ^b	
No.	R ₁	S.a.	<i>E.c.</i>	К.р.	M.m.	S.m.	po, 50 mg/kg, rats $(n=3)$	
<u>1</u> a	CH ₃	50	1.56	0.20	0.20	1.56	<0.05	
1b	CH ₂ COONa	>100	0.78	0.39	≦0.10	0.39	15.8	
1c	(CH ₂) ₂ COONa	>100	1.56	≦0.10	0.39	6.25	<0.4	
1d	CH(CH ₃)COONa	>100	1.56	≦0.10	0.20	1.56	0.9	
1e	C(CH ₃) ₂ COONa	100	1.56	≦0.10	0.20	0.39	<0.3	
1f	CH ₂ COOEt	50	12.5	0.78	1.56	6.25	<2.2	
1g	CH_2CONH_2	100	1.56	≤ 0.10	0.39	1.56	<2.1	
Cepl	nalexin	0.78	12.5	3.13	>100	>100	16.6	
Cefiz	kime°	25	0.78	≦0.10	0.20	0.78	30.2	

^a The MICs are determined by a standard agar dilution method using sensitivity test agar (Eiken, Japan).

^b The peak serum levels were measured by a disc-plate method using *Escherichia coli* SC 507 or *Micrococcus luteus* NIHJ as test organism.

This reference compound was prepared according to the reported procedure⁷).
 Abbreviations: S.a.; Staphylococcus aureus 209P JC-1, E.c.; Escherichia coli NIHJ JC-2, K.p.;

Klebsiella pneumoniae IFO 3317, M.m.; Morganella morganii IID 602, S.m.; Serratia marcescens IID 618.

Table 2. In vitro antibacterial activity and peak serum level of $1h \sim 11$.



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	Compound		MIC (µ	ug/ml, 10 ⁶ c	fu/ml)¤		Peak serum level	
No.	R ₂	S.a.	<i>E.c.</i>	K.p.	M.m.	S.m.	po, 50 mg/kg, rats $(n=3)$	
1b	Et	>100	0.78	0.39	≦0.10	0.39	15.8	
1h	Na	>100	12.5	1.56	6.25	50	<2.9	
1i	CH_3	>100	1.56	≤ 0.10	0.20	1.56	24.4	
11	iso-Pr	> 100	3.13	≦0.10	0.20	0.39	16.4	
1k	CH ₂ CH=CH ₂	>100	0.78	≦0.10	≦0.10	1.56	7.5	
11	CH ₉ Ph	>100	12.5	0.39	0.78	6.25	<3.1	
Cepha	alexin	0.78	12.5	3.13	>100	>100	16.6	
Cefixi	me°	25	0.78	≦0.10	0.20	0.78	30.2	

^{a~c} and organism abbreviations: See the footnote in Table 1.

and cefixime. Regarding the oral absorption of these compounds, only 1b with a carboxymethyl group as R_1 showed good oral absorption. Its peak serum level was comparable to that of cephalexin, but was lower than that of cefixime.

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Table 2 shows the influence of the C-3 substituent variation on 1b. Against the Gram-negative bacteria, the cephalosporins (1i, 1j and 1k) having a methyl, isopropyl or allyl ester group, respectively, in the C-3 substituent exhibited similar activity to that of 1b and cefixime. However, the derivatives (1h and 1l) bearing a carboxy function or benzyl ester group, respectively, were much less active. Against *S. aureus*, all of these derivatives displayed no significant activity, probably due to their high hydrophilicity. Regarding the oral absorption, compound 1i ($R_2=CH_3$) showed the best oral absorption in this series, but its peak serum level did not reach the level of cefixime. In contrast, the other analogues such as 1h, 1k and 1l exhibited low oral absorption.

The results shown in Table 2 indicate that the presence of the lower alkoxycarbonylmethoxy substituent at the C-3 position is of importance on antibacterial activity as well as oral absorption of these compounds.

In this study, we could improve the oral absorption in rats of cephalosporins carrying an 2-aminothiazole-oxime side chain by application of novel C-3 substituents which were found in a previous study on cephalexin analogues. However, further improvement of antibacterial activity against Gram-positive bacteria is desirable.

Experimental

MP was determined with a Yanagimoto micro melting point apparatus and are uncorrected. IR spectra were taken on a Jasco DS-701G IR spectrometer. ¹H NMR spectra were recorded on a Varian XL-200 NMR spectrometer using TMS or sodium trimethylsilyl propionate- d_4 (in D₂O) as an internal standard. MS was measured on a Jeol JMS-DX303 mass spectrometer. The following abbreviations are used: s, singlet; d, doublet; t, triplet; m, multiplet; br s, broad singlet; ABq, AB quartet.

Determination of Antibacterial Activity

MIC was determined by the agar dilution method using sensitivity test agar (Eiken, Japan) after incubation at 37° C for 18 hours with inoculum size of 10^{6} cfu/ml.

Oral Absorption Study

Male SLC/Wistar rats (n=3) weighing $180 \sim 220$ g were fasted overnight and orally dosed with 50 mg/kg of the test compounds. Serum samples were collected at 0.5, 1, 2 and 4 hours respectively after dosing. The serum levels of the test compounds were measured by the disc-plate method using *Escherichia coli* SC 507 or *Micrococcus luteus* NIHJ as the test organism and the sensitivity test agar (Eiken, Japan) as the test medium.

<u>Diphenylmethyl</u> 7β -Phenoxyacetamido-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate 1β -Oxide (4c)

To a solution of 2 (5.32 g, 10 mM) in DMSO (50 ml) were added ethyl bromoacetate (3.54 g, 21.2 mM) and *N*,*N*-diisopropylethylamine (1.94 g, 15 mM) at room temp. After being stirred at 45°C for 1 hour, the reaction mixture was poured into 0.5% HCl (80 ml) under ice-cooling and extracted with EtOAc (150 ml). The extract was washed with brine (100 ml), dried (MgSO₄) and evaporated. The residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 5:1), and crystallized from MeOH to give 2.50 g (40.5%) of 4c: MP 102~104°C. IR (KBr) cm⁻¹ 1780, 1730, 1690; ¹H NMR (CDCl₃) δ 1.24 (3H, t, J=7 Hz, CH₂CH₃), 3.49 (1H, dd, J=1.5 and 17 Hz, 2-H_a), 3.94 (1H, d, J=17 Hz, 2-H_β), 4.16 (2H, q, J=7 Hz, CH₂CH₃), 4.38 and 4.52 (2H, ABq, J= 16 Hz, OCH₂CO), 4.54 (1H, dd, J=1.5 and 5 Hz, 6-H), 4.57 (2H, s, PhOCH₂), 6.10 (1H, dd, J=5 and 9 Hz, 7-H), 6.90~7.08 (4H, m, CHPh₂ and aromatic H), 7.26~7.50 (12H, m, aromatic H), 7.89 (1H, d, J=9 Hz, CONH); field desorption mass spectrum (FD-MS) m/z 618 (M)⁺;

Anal Calcd for C₃₂H₃₀N₂O₉S: C 62.13, H 4.89, N 4.53.

Found: C 62.05, H 4.66, N 4.31.

Table 3. ¹H NMR, MS and IR spectral data and yield of 4a, 4b and $4d \sim 4f$.



(Compound				MR, δ (CDCl ₃)				
No.	R ₂	$\begin{array}{c} 2-H_{\alpha} \\ (1H, dd, \\ J=1.5, \\ 17 \text{ Hz}) \end{array}$	$2-H_{\beta}$ (1H, d, J=17 Hz)	$\begin{array}{c} 6-H \\ (1H, dd, \\ J=1.5, \\ 5 \text{ Hz}) \end{array}$	7-H (1H, dd, J=5, 9 Hz)	Other protons	MS (<i>m</i> / <i>z</i>)	IR (KBr) cm ⁻¹	Yield (%)
4 a	Bh	3.37	3.84	4.40	6.07	4.48 and 4.62 (2H, ABq, $J=16$ Hz), 4.56 (2H, s), 6.88 (1H, s), 6.90~7.08 (4H, m), 7.22~7.51 (22H, m), 7.87 (1H, d, $J=9$ Hz)	757 (M+1)+	1780, 1720	32
4b	CH ₃	3.48	3.94	4.55	6.11	3.70 (3H, s), 4.40 and 4.55 (2H, ABq, $J=16$ Hz), 4.59 (2H, s), 6.90~7.08 (4H, m), 7.24~7.52 (12H, m), 7.89 (1H, d, $J=9$ Hz)	604 (M)+	1780, 1750	38
4d	iso-Pr	3.49	3.94	4.54	6.09	1.21 (6H, d, $J=7$ Hz), 4.34 and 4.48 (2H, ABq, $J=17$ Hz), 4.57 (2H, s), 5.02 (1H, m), 6.90 \sim 7.07 (4H, m), 7.24 \sim 7.52 (12H, m), 7.88 (1H, d, $J=9$ Hz)	632 (M)+	1780, 1720	36
4 e	CH ₂ CH=CH ₂	3.49	3.95	4.56	6.11	4.42 and 4.57 (2H, ABq, $J=16$ Hz), 4.49~4.66 (2H, m), 4.58 (2H, s), 5.24~5.37 (2H, m), 5.76~ 5.98 (1H, m), 6.90~7.08 (4H, m), 7.24~7.52 (12H, m), 7.89 (1H, d, $J=9$ Hz)	630 (M)+	1780, 1750	39
4f	CH₂Ph	3.46	3.93	4.49	6.11	4.44 and 4.60 (2H, ABq, J=16 Hz), 4.60 (2H, s), 5.14 (2H, s), 6.90~7.10 (4H, m), 7.26~7.53 (17H, m), 7.90 (1H, d, J=9 Hz)	680 (M)+	1780, 1745, 1720	33

Table 4. ¹H NMR, MS and IR spectral data and yield of 5a, 5b and 5d~5f.

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(Compound				¹ H NMI	R, δ (CDCl ₃)		TD	
No.	R ₂	$2-H_{\alpha}$ (1H, d, J=17 Hz)	$2-H_{\beta}$ (1H, d, J=17 Hz)	6-H (1H, d, <i>J</i> =5 Hz)	7-H (1H, dd, J=5, 9 Hz)	Other protons		(KBr) cm ⁻¹	Yield (%)
5a	Bh	3.17	3.30	4.98	5.67	4.58 (4H, br s), 6.89~7.08 (5H, m), 7.20~7.46	740	1770,	95
						(22H, m), 7.52 (1H, d, J=9 Hz)	(M)+	1760	
5b	CH_3	3.13	3.35	5.07	5.66	3.72 (3H, s), 4.44 and 4.56 (2H, ABq, J=17 Hz),	588	1770,	85
						4.60 (2H, s), 6.90~7.08 (4H, m), 7.24~7.48	(M)+	1760	
						(12H, m), 7.64 (1H, d, J=9 Hz)			
5d	iso-Pr	3.28	3.38	5.09	5.69	1.23 (6H, d, $J=7$ Hz), 4.44 and 4.52 (2H, ABq,	616	1775,	88
						J=16 Hz), 4.60 (2H, s), 5.06 (1H, m), 6.92~7.10	(M)+	1760	
						(4H, m), 7.26~7.56 (13H, m)			
5e	$CH_2CH=CH_2$	3.17	3.36	5.07	5.67	4.42~4.66 (4H, m), 4.60 (2H, s), 5.24~5.39 (2H,	614	1770,	94
						m), 5.79~5.99 (1H, m), 6.90~7.08 (4H, m),	(M)+	1760	
						$7.24 \sim 7.48$ (12H, m), 7.62 (1H, d, $J=9$ Hz)			
5 f	CH_2Ph	3.24	3.35	5.02	5.69	4.29 and 4.59 (2H, ABq, J=17 Hz), 4.60 (2H, s),	664	1770,	86
						5.16 (2H, s), 6.92~7.08 (4H, m), 7.24~7.45	(M)+	1760	
						(17H, m), 7.51 (1H, d, J=9 Hz)			

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Table 5. ¹H NMR, MS and IR spectral data and yield of 6a, 6b and 6d~6f.



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(Compound				MR, δ (CDCl ₃)		TD			
No.	R ₂	$2-H_{\alpha}$ (1H, d, J=17 Hz)	$2-H_{\beta}$ (1H, d, J=17 Hz)	6-H (1H, d, J=5 Hz)	7-H (1H, d, J=5 Hz)	Other protons	(m/z)	(KBr) cm ⁻¹	Y ield (%)	
6a	Bh	3.34	3.54	4.67	4.88	1.75 (2H, br s), 4.56 (2H, s), 6.94 (1H, s), 6.96	607	1780	55	
						$(1H, s), 7.24 \sim 7.48 (20H, m)$	(M+1) ⁺			
6b	CH_3	3.39	3.60	4.69	4.95	1.82 (2H, br s), 3.71 (3H, s), 4.47 (2H, br s), 6.96	455	1760	56	
						(1H, s), 7.24~7.48 (10H, m)	(M+1)+			
6d	iso-Pr	3.40	3.59	4.69	4.97	1.22 (6H, d, J=7 Hz), 1.82 (2H, br s), 4.44 (2H, s),	482	1760	52	
						5.06 (1H, m), 6.96 (1H, s), 7.22~7.50 (10H, m)	(M)+			
6e	$CH_2CH = CH_2$	3.40	3.60	4.69	4.96	1.85 (2H, br s), 4.49 (2H, s), 4.58~4.65 (2H, m),	481	1760	47	
						5.24~5.39 (2H, m), 5.78~5.99 (1H, m), 6.97 (1H,	(M+1)+			
						s), 7.23~7.50 (10H, m)				
6f	CH₂Ph	3.37	3.58	4.69	4.92	1.75 (2H, br s), 4.50 (2H, s), 5.15 (2H, s), 6.96	531	1760	56	
	(1H, s					(1H, s), 7.26~7.50 (15H, m)	(M+1)*			

Table 6. ¹H NMR, MS and IR spectral data and yield of 8a and 8c~8l.



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Compound					¹ H NMR	, δ (CDCl ₃)		at	
No.	R ₁	R₂	$ \begin{array}{c} 6-H \\ (1H, d, \\ J=5 \text{ Hz}) \end{array} $	7-H (1H, dd, J=5, 9 Hz)	Thiazole 5-H (1H, s)	Other protons	MS (m/z)	(KBr) cm ⁻¹	Yield (%)
8a	CH ₃	Et	5.11	5.76	6.80	1.25 (3H, t, J=7 Hz), 3.38 (2H, br s), 4.08 (3H, s), 4.19 (2H, q, J=7 Hz), 4.50 (2H, br s), 6.93 (1H, s), 7.02 (1H, br s), 7.20~7.46 (26H, m)	893 (M) ⁺	1775, 1720	70
80	(CH₂)₂COOBh	Et	4.98	5.57	6.78	1.24 (3H, t, $J=7$ Hz), 2.90 (2H, t, $J=6$ Hz), 3.28 (2H, br s), 4.18 (2H, q, $J=7$ Hz), 4.26 (2H, br s), 4.62 (2H, t, $J=6$ Hz), 6.88 (1H, s), 6.94 (1H, s), 6.98 (1H, br s), 7.08 (1H, d, $J=9$ Hz), 7.20~7.48 (35H, m)	1,117 (M) ⁺	1780, 1725	43
8d	CH(CH₃)COOBh	Et	5.06	5.76 and 5.86	6.81 and 6.82	1.24 and 1.26 (3H, t, $J=7$ Hz), 2.82~3.16 (2H, m), 4.17 and 4.20 (2H, q, $J=7$ Hz), 4.49 and 4.55 (2H, br s), 5.15~5.30 (1H, m), 6.85 and 6.86 (1H, s), 6.92 (1H, s), 7.00 (1H, br s), 7.10~7.48 (35H, m), 8.27 (1H, d, $J=9$ Hz)	1,118 (M+1)+	1780, 1730	45
8e	C(CH ₃) ₂ COOBh	Et	5.04	5.84	6.72	1.25 (3H, t, $J=7$ Hz), 1.71 (3H, s), 1.74 (3H, s), 3.12 (1H, d, $J=17$ Hz), 3.22 (1H, d, $J=17$ Hz), 4.19 (2H, q, $J=7$ Hz), 4.51 (2H, s), 6.86 (1H, s), 6.94 (1H, br s), 6.96 (1H, s), 7.20~7.53 (36H, m)	1,131 (M) ⁺	1780, 1725	50

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8f	CH ₂ COOEt	Et	5.12	5.77	6.87	1.25 (3H, t, <i>J</i> =7 Hz), 1.27 (3H, t, <i>J</i> =7 Hz), 3.42 (2H, br s), 4.18 (2H, q, <i>J</i> =7 Hz), 4.21 (2H, q, <i>J</i> =7 Hz), 4.56 (2H, br s), 4.88 (2H, br s), 6.92 (1H, s), 7.00 (1H, br s) 7.20~7.47 (25H, m), 8.47 (1H, d, <i>J</i> =9 Hz)	965 (M)+	1780, 1725	60
8 g	CH ₂ CONH ₂	Et	4.98	5.75	6.80	1.26 (3H, t, $J=7$ Hz), 1.80 (2H, br s), 3.24 (2H, br s), 4.19 (2H, q, $J=7$ Hz), 4.50 (2H, br s), 4.56 and 4.68 (2H, ABq, $J=$ 16 Hz), 6.90 (1H, s), 7.10~7.40 (27H, m)	937 (M+1)+	1780, 1720	64ª
8h	CH₂COOBh	Bh	4.95	5.75	6.83	2.94 (1H, d, $J=17$ Hz), 3.02 (1H, d, $J=$ 17 Hz), 4.58 (2H, s), 4.92 and 5.05 (2H, ABq, $J=17$ Hz), 5.90 (2H, s), 5.92 (1H, s), 7.00 (1H, br s), 7.16~7.46 (45H, m), 8.19 (1H, d, $J=9$ Hz)	1,242 (M+1)+	1775, 1730	49
8i	CH₂COOBh	$ m CH_3$	5.06	5.77	6.85	3.03 (1H, d, $J=17$ Hz), 3.13 (1H, d, $J=17$ Hz), 3.71 (3H, s), 4.51 (2H, s), 4.94 and 5.07 (2H, ABq, $J=17$ Hz), 6.93 (1H, s), 6.94 (1H, s), 7.00 (1H, br s), 7.24~ 7.48 (35H, m), 8.20 (1H, d, $J=9$ Hz)	1,090 (M+1)+	1775, 1735	44
8j	CH₂COOBh	iso-Pr	5.05	5.75	6.84	1.23 (6H, d, $J=7$ Hz), 3.01 (1H, d, $J=17$ Hz), 3.10 (1H, d, $J=17$ Hz), 3.10 (1H, d, $J=17$ Hz), 4.46 (2H, s), 4.94 and 5.05 (2H, ABq, $J=17$ Hz), 5.02 (1H, m), 6.91 (1H, s), 6.92 (1H, s), 7.00 (1H, br s), 7.20~7.48 (35H, m), 8.17 (1H, d, $J=9$ Hz)	1,118 (M+1)+	1775, 1725	56
8k	CH₃COOBh	CH ₂ CH=CH ₂	5.05	5.77	6.84	3.03 (1H, d, $J=17$ Hz), 3.12 (1H, d, $J=$ 17 Hz), 4.52 (2H, s), 4.61 (2H, d, $J=6$ Hz), 4.94 and 5.06 (2H, ABq, $J=17$ Hz), 5.20~ 5.38 (2H, m), 5.77~5.99 (1H, m), 6.92 (1H, s), 6.94 (1H, s), 7.01 (1H, br s), 7.20~7.48 (35H m) 8.19 (1H d, $J=9$ Hz)	1,116 (M+1)+	1770, 1730	48
81	CH₂COOBh	CH ₂ Ph	4.99	5.75	6.84	2.97 (1H, d, $J=17$ Hz), 3.06 (1H, d, $J=$ 17 Hz), 4.53 (2H, s), 4.94 and 5.05 (2H, ABq, $J=16$ Hz), 5.14 (2H, s), 6.91 (1H, s), 6.92 (1H, s), 7.00 (1H, br s), 7.22~7.46 (40H, m), 8.18 (1H, d, $J=9$ Hz)	1,166 (M+1)*	1775, 1730	55

* This compound was prepared from 6c and 7 ($R_1 = CH_2CONH_2$) by using N,N'-dicyclohexylcarbodiimide as a condensation reagent.

Tr: CPh₃.

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The spectral data and yield of various derivatives (4) are listed in Table 3.

Diphenylmethyl 7β -Phenoxyacetamido-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate (5c)

To a solution of 4c (2.85 g, 4.61 mM) in DMF (25 ml) was added dropwise phosphorus tribromide (1.25 g, 4.61 mM) under ice-cooling. After being stirred at the same temp for 30 minutes, the reaction mixture was poured into H₂O (100 ml) and extracted with EtOAc (100 ml). The extract was washed with brine (50 ml), dried (MgSO₄) and evaporated. The residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 20:1) to give 2.30 g (83%) of 5c as an amorphous solid: IR (KBr) cm⁻¹ 1760, 1680; ¹H NMR (CDCl₃) δ 1.26 (3H, t, J=7 Hz, CH₂CH₃), 3.29 (1H, d, J=17 Hz, 2-H_a), 3.40 (1H, d, J=17 Hz, 2-H_β), 4.19 (2H, q, J=7 Hz, CH₂CH₃), 4.46 and 4.54 (2H, ABq, J=17 Hz, OCH₂CO), 4.59 (2H, s, PhOCH₂), 5.08 (1H, d, J=5 Hz, 6-H), 5.71 (1H, dd, J=5 and 9 Hz, 7-H), 6.90~7.10 (4H, m, CHPh₂ and aromatic H), 7.26~7.52 (13H, m, CONH and aromatic H); FD-MS m/z 602 (M)⁺.

The spectral data and yield of various derivatives (5) are listed in Table 4.

Diphenylmethyl 7β -Amino-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate (6c)

To a solution of 5c (2.15 g, 3.57 mM) in CH₂Cl₂ (40 ml) were added pyridine (0.85 g, 10.8 mM) and phosphorus pentachloride (1.49 g, 7.15 mM) at -30° C. The temp of the mixture was raised to room temp over 30 minutes. After being stirred at the same temp for 1 hour, the reaction mixture was cooled to -50° C. To the cooled mixture was added MeOH (20 ml) all at once, and then stirred for 1 hour under ice-cooling. Subsequently, the reaction mixture was cooled to -30° C, and H₂O (20 ml) was added. After being stirred for 45 minutes under ice-cooling, organic solvents were removed under reduced pressure, maintaining the temp below 10°C. The resulting aqueous solution was neutralized with 5% aq NaHCO₃ and extracted with CH₂Cl₂ (100 ml). The extract was washed with brine (50 ml), dried (MgSO₄) and evaporated. The residue was purified by column chromatography on silica gel (eluent; benzene - EtOAc, 1:1), and crystallized from Et₂O to give 1.0 g (60%) of **6c**: MP 94~96°C; IR (KBr) cm⁻¹ 1765, 1750, 1720; ¹H NMR (CDCl₃) δ 1.24 (3H, t, J=7 Hz, CH₂CH₃), 1.80 (2H, br s, NH₂), 3.39 (1H, d, J=17 Hz, 2-H_a), 3.59 (1H, d, J=17 Hz, 2-H_β), 4.17 (2H, q, J=7Hz, CH₂CH₃), 4.45 (2H, s, OCH₂CO), 4.68 (1H, d, J=5 Hz, 6-H), 4.95 (1H, d, J=5 Hz, 7-H), 6.95 (1H, s, CHPh₂), 7.26~7.50 (10H, m, aromatic H); FD-MS m/z 468 (M)⁺;

Anal Calcd for $C_{24}H_{24}N_2O_6S$: C 61.53, H 5.16, N 5.98.

Found: C 61.54, H 5.12, N 5.93.

The spectral data and yield of various derivatives (6) are listed in Table 5.

<u>Diphenylmethyl</u> 7β -[2-(2-Tritylaminothiazol-4-yl)-2-[(Z)-diphenylmethoxycarbonylmethoxyimino]acetamido]-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate (**8b**)

To a solution of 7 (R_1 =CH₂COOBh) (627 mg, 0.96 mM) in CH₂Cl₂ (18 ml) were added pyridine (380 mg, 4.80 mM) and phosphorus pentachloride (201 mg, 0.96 mM) at -15° C, and the mixture was stirred at $-15 \sim -10^{\circ}$ C for 15 minutes. Then, the solution of **6c** (300 mg, 0.64 mM) in CH₂Cl₂ (3 ml) was added to the above mixture at -20° C. After being stirred at $-20 \sim -10^{\circ}$ C for 20 minutes, 0.5% HCl (20 ml) was added to the reaction mixture, and extracted with EtOAc (100 ml). The extract was washed with brine (50 ml), dried (MgSO₄) and evaporated. The residue was purified by column chromatography on silica gel (eluent; benzene - EtOAc, 5:1) to give 353 mg (50%) of **8b** as an amorphous solid: IR (KBr) cm⁻¹ 1780, 1730, 1680; ¹H NMR (CDCl₃) δ 1.26 (3H, t, J=7 Hz, CH₂CH₃), 3.04 (1H, d, J=16 Hz, 2-H_a), 3.12 (1H, d, J=16 Hz, 2-H_β), 4.19 (2H, q, J=7 Hz, CH₂CH₃), 4.51 (2H, s, OCH₂CO), 4.59 and 5.06 (2H, ABq, J=17 Hz, NOCH₂CO), 5.07 (1H, d, J=5 Hz, 6-H), 5.77 (1H, dd, J=5 and 9 Hz, 7-H), 6.86 (1H, s, thiazole 5-H), 6.93 (1H, s, CHPh₂), 6.94 (1H, s, CHPh₂), 7.02 (1H, br s, Ph₃CNH), 7.20~7.49 (35H, m, aromatic H), 8.19 (1H, d, J=9 Hz, CONH); FD-MS m/z 1,104 (M+1)⁺.

The spectral data and yield of various derivatives (8) are listed in Table 6.

Sodium 7β -[2-(2-Aminothiazol-4-yl)-2-[(Z)-carboxymethoxyimino]acetamido]-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate (1b)

To a mixture of TFA (4 ml) and anisole (0.8 ml) was added 8b (300 mg, 0.27 mM) under ice-cool-

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Table 7. ¹H NMR and IR spectral data and yield of 1a and $1c \sim 11$.



						1				
	Compound	<u> </u>				¹ H NM	$\overline{\mathbf{R}, \delta (\mathbf{D}_2 \mathbf{O})}$		ŤΒ	
No.	\mathbf{R}_1	R ₂	$\begin{array}{c} 2-H_{\alpha} \\ (1H, d, \\ J=17 \text{ Hz}) \end{array}$	$2-H_{\beta}$ (1H, d, J=17 Hz)	6-H (1H, d, J=5 Hz)	7-H (1H, d, J=5 Hz)	Thiazole 5-H (1H, s)	Other protons	(KBr) cm ⁻¹	Yield (%)
1a	CH ₃	Et	3.44	3.72	5.24	5.71	7.08	1.29 (3H, t, $J=7$ Hz), 4.01 (3H, s), 4.28 (2H, q, $J=7$ Hz), 4.61 and 4.72 (2H, ABq, $J=16$ Hz)	1755	95
1c	(CH ₂) ₂ COONa	Et	3.45	3.71	5.24	5.70	7.08	1.29 (3H, t, $J=7$ Hz), 2.64 (2H, t, J=7 Hz), 4.28 (2H, q, $J=7$ Hz), 4.44 (2H, t, $J=7$ Hz), 4.61 and 4.73 (2H, ABq, $J=16$ Hz)	1750	90
1d	CH(CH ₃)COONa	Et	3.44 and 3.46	3.69	5.26	5.72 and 5.75	7.08	1.28 (3H, t, $J=7$ Hz), 1.47 and 1.48 (3H, d, $J=7$ Hz), 4.27 (2H, q, $J=7$ Hz), 4.62 and 4.73 (2H, ABq, J=16 Hz)	1750	89
1e	C(CH ₃) ₂ COONa	Et	3.44	3.70	5.25	5.73	7.06	1.28 (3H, t, J=7 Hz), 1.50 (3H, s), 1.52 (3H, s), 4.27 (2H, q, J=7 Hz), 4.61 and 4.73 (2H, ABq, J=17 Hz)	1755	88
1f	CH ₂ COOEt	Et	3.43	3.71	5.25	5.74	7.15	1.28 (6H, t, $J=7$ Hz), 4.28 (2H, q, J=7 Hz), 4.29 (2H, q, $J=7$ Hz), 4.61 and 4.72 (2H, ABq, $J=17$ Hz), 4.85 (2H, s)	1750	91
1g	CH ₂ CONH ₂	Et	3.44	3.72	5.26	5.75	7.18	1.29 (3H, t, $J=7$ Hz), 4.28 (2H, q, J=7 Hz), 4.62 and 4.72 (2H, ABq, J=17 Hz), 4.77 (2H, s)	1750	87
1h	CH₂COONa	Na	3.40	3.62	5.28	5.69	7.15	4.36 and 4.48 (2H, ABq, $J=16$ Hz), 4.59 (2H, s)	1750	80

CH₂COONa

CH₂COONa

CH₂COONa

CH₂COONa

1i

1j

1k

11

 CH_3

iso-Pr

 $CH_2CH = CH_2$

 CH_2Ph

3.44

3.44

3.44

3.37

3.70

3.69

3.70

3.59

5.23

5.26

5.25

5.14

5.74

5.73

5.74

5.70

7.11

7.12

7.12

7.10

3.80 (3H, s), 4.60 (2H, s)

16 Hz), 5.12 (1H, m)

5.90~6.12 (1H, m)

(5H, br s)

1.29 (6H, d, J=7 Hz), 4.60 (2H,

br s), 4.61 and 4.72 (2H, ABq, J =

4.59 (2H, s), 5.26~5.46 (2H, m),

4.59 (2H, s), 5.29 (2H, s), 7.48

92

90

89

88

1750

1750

1750

1750

ing. After being stirred at the same temp for 40 minutes, the resulting solution was added dropwise to a mixture of Et_2O and *n*-hexane (1:2, 30 ml). The precipitated trifluoroacetate of the desired product was collected by filtration, and washed with a mixture of Et_2O and *n*-hexane (1:2, 40 ml). The above trifluoroacetate and NaHCO₃ (68 mg, 0.81 mM) were dissolved in H₂O (5 ml), and the solution was treated with column chromatography on Sephadex LH-20 (eluent; H₂O), and then lyophilized to give 140 mg (90%) of 1b as a white solid: IR (KBr) cm⁻¹ 1750; ¹H NMR (D₂O) δ 1.28 (3H, t, J= 7 Hz, CH₂CH₃), 3.43 (1H, d, J=17 Hz, 2-H_α), 3.70 (1H, d, J=17 Hz, 2-H_β), 4.28 (2H, q, J=7 Hz, CH₂CH₃), 4.60 (2H, s, NOCH₂CO), 5.27 (1H, d, J=5 Hz, 6-H), 5.74 (1H, d, J=5 Hz, 7-H), 7.12 (1H, s, thiazole 5-H).

The spectral data and yield of various derivatives (1) are listed in Table 7.

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